

Acid-Induced Rearrangement of Epoxygermacra-8,12-olides: Synthesis and Absolute Configuration of Guaiane and Eudesmane Derivatives from Artemisiifolin

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A study on the acid-induced rearrangement of 4,5-epoxy- and 1,10-epoxygermacra-8,12-olides was carried out. From a 4,5-epoxy derivative, guaianes were obtained, whereas 1,10-

epoxy derivatives furnished, depending on the stereochemistry of the C-1/C-10 epoxy ring, *trans*-5 β ,10 α - or *trans*-5 α ,10 β -eudesmanolides.

Introduction

Sesquiterpene lactones (SQLs) are an important class of plant secondary metabolites with antitumoural, phytotoxic, antimicrobial and other biological properties.^[1–3] Inside this class, germacranolides are considered to be the precursors of the other skeletal types of SQLs such as elemanolides, eudesmanolides and guaianolides.^[1] Acid-induced cyclization of germacranolides or that of their corresponding monoepoxides is generally accepted to be biomimetic and represents the biosynthesis of eudesmane and guaiane sesquiterpene lactones.^[4–11] In the frame of our ongoing research^[12,13] aimed to synthesize natural compound derivatives with potential antitumour activity, some years ago we published a paper on the synthesis of elemene and heliangolane derivatives,^[14] and we recently reported on the reactivity of epoxy derivatives and their cytotoxic properties,^[15] all prepared starting from cnicin (**1**), which is a 6,12-germacranolide commonly found in species of *Centaurea*, whose antibacterial, cytotoxic and antifungal properties have been investigated.^[16–18] In that work, it was shown that acid treatment of the 1 β ,10 α -epoxide results in the formation of eudesmanolides, whereas when the 4 α ,5 β -epoxide was treated similarly, no transformation was obtained. As continuation of our studies on cyclization reactions of germacranolides, we investigated the acid-induced cyclization of the monoepoxides of artemisiifolin, a natural germacra-8,12-olide isolated from *Ambrosia artemisiifolia*^[19] and obtained by us from cnicin by basic hydrolysis. NMR spectra of artemisiifolin show broad or even additional signals, indicating the existence of four conformers in solution as described previously.^[20] A smooth epoxidation of one double bond in *E,E*-germacra-8,12-olides decreases the confor-

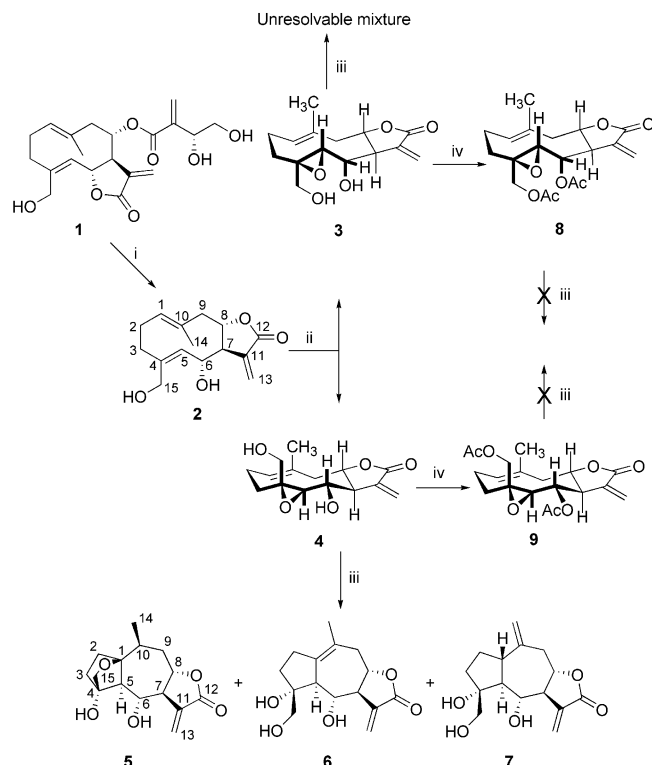
mational flexibility of the 10-membered ring, and the stereochemistry of the resulting epoxides should be affected by the relative abundance of the equilibrating conformers. The epoxy group can be described as a conformational “anchor”. Because the regiochemistry and stereochemistry of epoxides play a pivotal role in the outcome of the acid-induced cyclization, our aim was to rationalize the consequences of the conformational equilibrium existing for artemisiifolin on biomimetic cyclizations of its epoxides by experimental data. The lack of such data is pointed out in many theoretical work.^[21]

Results and Discussion

Cnicin (**1**) was isolated in large amounts from *Centaurea napifolia*,^[22] providing us with enough starting material to plan this study. Strong basic treatment^[23] of cnicin (**1**) resulted in the hydrolysis of the side chain and in relactonization of the C-8 position to afford artemisiifolin (**2**; Scheme 1). Epoxidation of compound **2** with *meta*-chloroperbenzoic acid (*m*-CIPBA) in pyridine gave 4,5-epoxides **3** and **4** in a 3:1 ratio. The major product was 4 β ,5 α -epoxyartemisiifolin (**3**), as clearly indicated by the 5-H/6-H coupling constant ($J = 3.3$ Hz) and by the chemical shifts of 6-H ($\delta_{\text{H}} = 4.12$ ppm) and 9 β -H ($\delta_{\text{H}} = 1.99$ ppm).^[24] Similar considerations allowed us to assign the structure of 4 α ,5 β -epoxyartemisiifolin to compound **4**. Compounds **3** and **4** arise from the epoxidation of the four conformers of artemisiifolin, and their stereochemistry and relative abundance are in perfect agreement with theoretical calculations.^[20]

Epoxide **4** was treated with HCl/CHCl₃ to yield guaianolides **5–7**. The ¹H NMR, ¹³C NMR and 2D spectra of compound **5** showed, apart from the signals of the lactone, the presence of an α -proton at $\delta_{\text{H}} = 1.88$ ppm (d, 5-H) coupling with 6-H ($J = 8.7$ Hz), two quaternary oxygenated carbon atoms at $\delta_{\text{C}} = 88.5$ and 83.0 ppm involved in the formation of a tetrahydrofuran ring (C-1, C-5, C-4, C-15) and a

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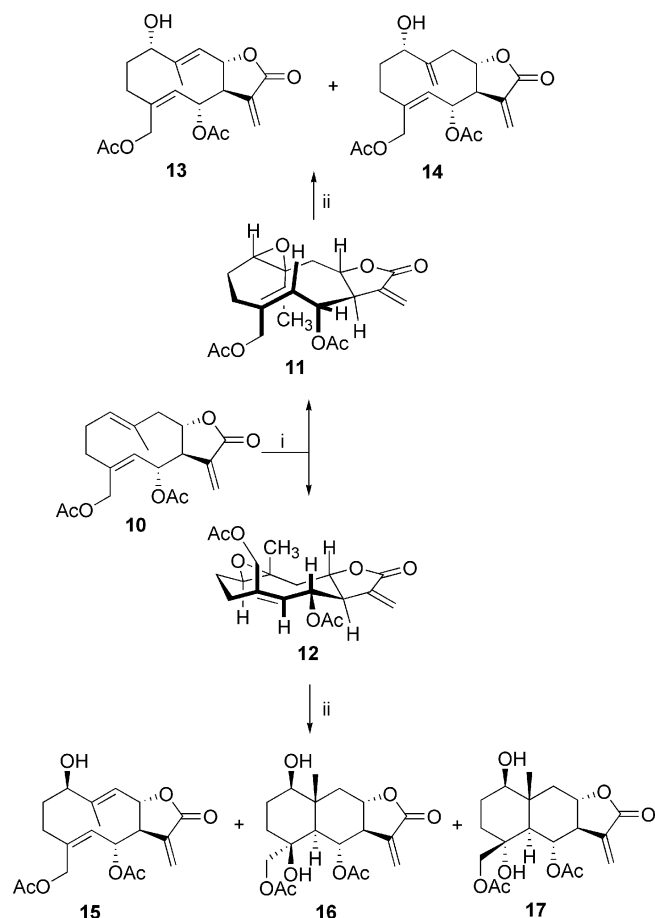


Scheme 1. Reagents and conditions: (i) KOH, H₂O, room temp., 4 h, 77%; (ii) *m*-CIPBA, THF, room temp., 24 h; (iii) CHCl₃, HCl/CHCl₃(sat.), room temp., 96 h; (iv) Ac₂O, pyridine, room temp., 2 h, 100%.

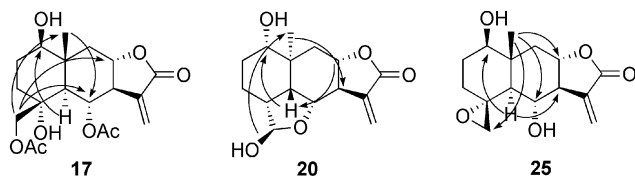
methyl group at $\delta_{\text{H}} = 1.15$ ppm (d, $J = 7.4$ Hz, 3 H, 14-H₃) coupling with 10-H, whose stereochemistry was assessed to be β as indicated by the coupling constants of 9-H^A and 9-H^B with 10-H. Compound 6, on the other hand, had instead of the tetrahydrofuran ring a hydroxymethyl group ($\delta_{\text{H}} = 3.93$ ppm, d, $J = 11.1$ Hz, 15-H_A; $\delta_{\text{H}} = 3.63$ ppm, d, $J = 11.1$ Hz, 15-H_B; $\delta_{\text{C}} = 64.2$ ppm, t, C-15) and a tetrasubstituted double bond ($\delta_{\text{C}} = 126.3$ ppm, s, C-1; $\delta_{\text{C}} = 131.6$ ppm, s, C-10). The spectroscopic data of compound 7 indicate that it was a double-bond isomer of triol 6. In fact, in this case, there were signals for an exocyclic double bond ($\delta_{\text{C}} = 144.7$ ppm, s, C-10; $\delta_{\text{C}} = 112.4$ ppm, t, C-14) and another allylic proton ($\delta_{\text{H}} = 2.17$ ppm, 1-H). The stereochemistry of 1-H was assigned to β by the value of the coupling constant with 5-H ($J = 11.0$).^[25,26] On the other hand, acidic treatment of compound 3 gave an irresolvable, complex mixture of products. Acetylation of compounds 3 and 4 furnished diacetyl epoxides 8 and 9, respectively, and when these were treated with HCl/CHCl₃, they were recovered unaltered.

The presence of two free hydroxy groups in artemisiifolin (2) induced selective epoxidation of the C-4/C-5 double bond. Consequently, we tried to direct the epoxidation on the other double bond (C-1/C-10) by transforming artemisiifolin (2) into diacetyl derivative 10.^[27] Therefore, by epoxidation of diacetyl artemisiifolin (10) only diastereoisomeric 1,10-epoxides 11 and 12 (15-acetoxypyrethrosin),^[28] in a 1:1 ratio, were obtained (Scheme 2). The stereochemistry of the

epoxy ring in compounds 11 and 12 was easily deduced by the chemical shifts of 14-H₃, 7-H and the two protons on C-9.^[24] Acid treatment of 1 α ,10 β -epoxide 11 gave the isomeric germacatrienes 13 and 14. The difference between the two compounds was the position of the double bond: C-9/C-10 in the former ($\delta_{\text{C}} = 126.0$ ppm, d, C-9; $\delta_{\text{C}} = 142.8$ ppm, s, C-10) and C-10/C-14 ($\delta_{\text{C}} = 146.6$ ppm, s, C-10; $\delta_{\text{C}} = 115.8$ ppm, t, C-14) in the latter. The same reaction, performed on 1 β ,10 α -epoxide 12, afforded germacrane 15, the C-1 epimer of compound 13, and eudesmanolides 16 and 17. The structure of the most abundant one (i.e., 17) was elucidated by extensive 1D and 2D NMR experiments, and the stereochemistry of the stereogenic centres was determined by NOESY (Figure 1). In fact, the correlations of 6-H ($\delta_{\text{H}} = 5.65$ ppm) with 14-H₃ ($\delta_{\text{H}} = 1.06$ ppm) and 15-H^A ($\delta_{\text{H}} = 4.38$ ppm) and the correlation of 8-H ($\delta_{\text{H}} = 4.01$ ppm) with both 15-H protons ($\delta_{\text{H}} = 4.38$ and 4.35 ppm) indicated the β orientation of the methyl 14-H₃ group and the acetoxymethylene group at C-15, whereas the correlation between the α -axial proton 1-H ($\delta_{\text{H}} = 3.53$ ppm, dd, $J = 10.7$, 5.0 Hz) with 5-H ($\delta_{\text{H}} = 2.03$ ppm) showed the *trans* fusion of the decalin moiety. In similar way, compound 16 was showed to be the C-4 epimer of eudesmanolide 17.

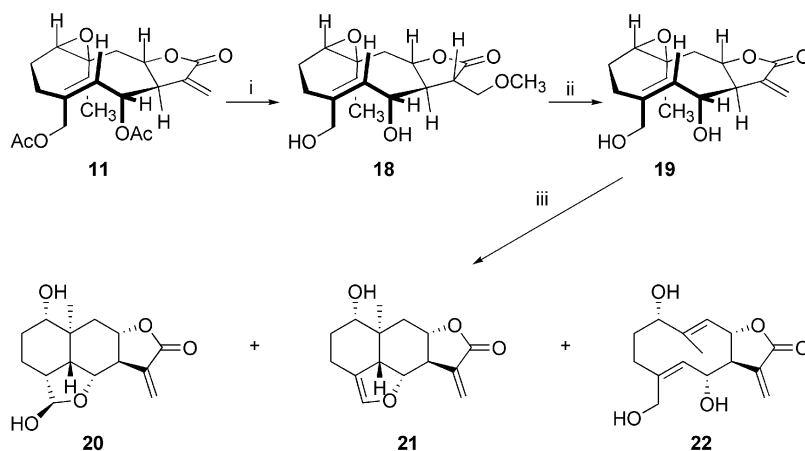
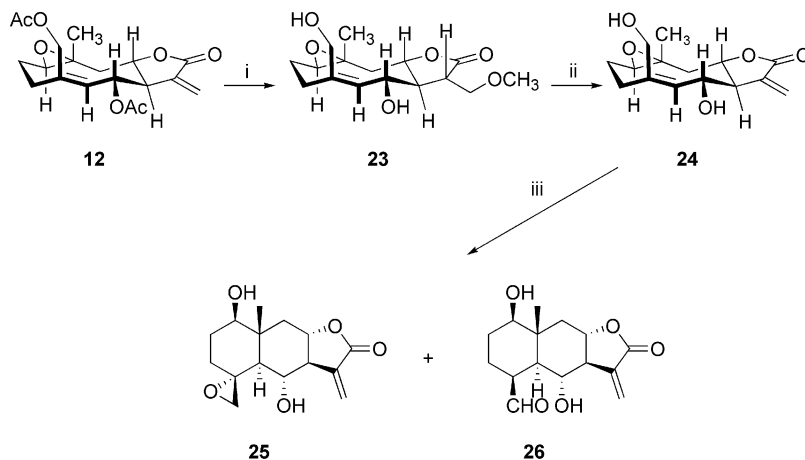


Scheme 2. Reagents and conditions: (i) *m*-CIPBA, THF, room temp., 48 h; (ii) CHCl₃, HCl/CHCl₃ (sat.), room temp., 24 h.

Figure 1. NOESY correlations of compounds **17**, **20** and **25**.

In order to exploit the influence of the acetoxy groups in the cyclization reaction of 1,10-epoxygermacranolides, we tried to hydrolyze the ester moieties of compound **11** by treating it with CH_3OH and NaHCO_3 . The reaction resulted in the removal of the two acetyl groups but also in the addition of a methoxy group at C-13 to afford quantitatively compound **18** (Scheme 3). The restoration of the C-11/C-13 exocyclic double bond was easily achieved by treating compound **18** with $t\text{BuOK}$ to give diol **19**. Reaction of compound **19** with HCl yielded eudesmanolides **20** and **21** and germacranolide **22**, which was the most abundant and which was identified as the deacetyl derivative of compound

13. Compound **20** showed signals for an axial proton at $\delta_{\text{H}} = 3.51$ ppm (dd, $J = 9.4, 5.6$ Hz, 1-H), an emiacetalic moiety at $\delta_{\text{H}} = 5.43$ ppm (d, $J = 6.0$ Hz, 15-H; $\delta_{\text{C}} = 104.8$ ppm, d, C-15) and an angular methine at $\delta_{\text{H}} = 2.05$ ppm (dd, $J = 5.5, 2.2$ Hz, 5-H) coupling with 4-H at $\delta_{\text{H}} = 2.26$ ppm and 6-H at $\delta_{\text{H}} = 4.57$ ppm. The stereochemistry of the stereogenic centres was determined by NOESY (Figure 1). In fact, the correlation of 14- H_3 ($\delta_{\text{H}} = 1.27$ ppm) with 7-H ($\delta_{\text{H}} = 3.38$ ppm) indicated the α orientation of the methyl 14- H_3 group, whereas the correlation between 5-H ($\delta_{\text{H}} = 2.05$ ppm) with the β -axial proton 1-H ($\delta_{\text{H}} = 3.51$ ppm) and 8-H ($\delta_{\text{H}} = 4.09$ ppm) showed the β orientation of 5-H and the $5\beta,10\alpha$ -trans fusion of the decalin moiety. Finally, the correlation between 15-H ($\delta_{\text{H}} = 5.43$ ppm) and 14- H_3 ($\delta_{\text{H}} = 1.27$ ppm) assessed an α stereochemistry for the emiacetalic proton. Compound **21** was the dehydration derivative of **20** as easily recognized by the absence of the emiacetalic signals and the presence of a trisubstituted double bond ($\delta_{\text{H}} = 6.18$ ppm, m, 15-H; $\delta_{\text{C}} = 112.6$ ppm, s, C-4; $\delta_{\text{C}} = 138.1$ ppm, d, C-15) involved in a dihydrofuran moiety.

Scheme 3. Reagents and conditions: (i) NaHCO_3 , CH_3OH , room temp., 24 h, 90%; (ii) $t\text{BuOK}$, THF, room temp., 10 min, 56%; (iii) CHCl_3 , HCl/CHCl_3 (sat.), room temp., 24 h.Scheme 4. Reagents and conditions: (i) NaHCO_3 , CH_3OH , room temp., 24 h, 96%; (ii) $t\text{BuOK}$, THF, room temp., 10 min, 61%; (iii) CHCl_3 , HCl/CHCl_3 (sat.), room temp., 24 h.

Also diacetyl 1 β ,10 α -epoxide **12** was subjected to basic hydrolytic conditions to afford methoxy derivative **23** (Scheme 4), which was easily transformed (*t*BuOK) into diol **24**. The acidic rearrangement of compound **24** gave, in this case, only eudesmanolides **25** and **26**. The NMR spectra of compound **25** indicated the presence of an axial proton at $\delta_{\text{H}} = 3.59$ ppm (dd, $J = 10.5, 5.0$ Hz, 1-H), a *trans* diaxial relationship between 5-H ($\delta_{\text{H}} = 1.86$ ppm, d, $J = 9.6$ Hz) and 6-H ($\delta_{\text{H}} = 3.85$ ppm, dd, $J = 9.6, 9.6$ Hz) and the presence of an epoxide ($\delta_{\text{H}} = 3.20$ ppm, dd, $J = 3.6, 1.2$ Hz, 15-H_A; $\delta_{\text{H}} = 2.86$ ppm, d, $J = 3.6$ Hz, 15-H_B; $\delta_{\text{C}} = 60.9$ ppm, C-4; $\delta_{\text{C}} = 51.9$ ppm, C-15), α -orientated, as indicated by the long-range coupling constant ($J = 1.2$ Hz) between 15-H^A and H-3^A.^[29,30] NOESY experiments confirmed the stereochemistry of the chiral centres (Figure 1). In fact, 14-H₃ ($\delta_{\text{H}} = 1.00$ ppm) correlated with 6-H, 8-H ($\delta_{\text{H}} = 3.96$ ppm) and both 15-H protons, whereas the correlation between 5-H with the α -axial proton 1-H and 7-H ($\delta_{\text{H}} = 2.54$ ppm) showed the α orientation of 5-H and the *trans* fusion of the decalin moiety. Compound **26** had a similar framework with respect to epoxide **25**. The only differences were the absence of the epoxy ring and the presence of an aldehyde ($\delta_{\text{H}} = 9.85$ ppm, br. s, 15-H; $\delta_{\text{C}} = 204.7$ ppm). The β -orientation of the aldehyde group was evident by the very small coupling constant with 4-H (<1 Hz), as reported for other compounds with a similar moiety.^[7,31]

Finally, because the absolute stereochemistries of cnicin (**1**) and artemisiifolin (**2**) were determined by X-ray analysis,^[20,32] in this paper, the absolute configurations of compounds **3–26** were ascertained.

Conclusions

The studied acid-catalyzed cyclizations of epoxygermacrane follow the well-established rule that 1,10-epoxygermacranolides cyclize into eudesmanolides^[33,34] under acid catalysis, whereas 4,5-epoxygermacranolides yield guaionolides,^[35] but interesting information concerning the stereochemistry of the epoxides and the presence of substituents can be deduced.

In these cyclizations, it is extremely important to consider the conformational features of the germacranolide ring; in fact cyclization of 4 α ,5 β -epoxyinunolide^[36] produces *cis*-fused guaianes, whereas epoxide **4**, with analogous stereochemistry, gives among other products, guaiane **7**, with a *trans*-fused junction, because **4** arises from epoxidation of two conformers^[20] of artemisiifolin (**2**) that can give the 4 α ,5 β -epoxy group, and one of these conformers has the suitable geometry for the *trans* fusion observed.

It is worthy to note that 1 α ,10 β -epoxy germacranolide **11** does not cyclize into eudesmanolide. On the contrary 1 β ,10 α -epoxide **12** gives eudesmanolides. When acetoxy groups are removed from scaffold, the acid-catalyzed cyclization of 1 α ,10 β -epoxy germacranolide **19** produces eudesmanolides **20** and **21**, probably because the two acetoxy groups induce strain in the conformation of the germacrane

ring, forcing the two reactive carbon atoms (C-5 and C-10) apart. An additional aspect to rationalize the cyclization of diol **19** in contrast to diacetate **11** is the more nucleophilic 4,5 double bond in compound **19**.

Experimental Section

General: IR spectra were obtained with a Perkin–Elmer 1310 spectrometer. ¹H NMR spectra were recorded in CDCl₃ or CD₃OD solution by using a Bruker Avance series 300 MHz spectrometer, and chemical shifts are reported with respect to residual CHCl₃ ($\delta = 7.27$ ppm) or CD₃OD ($\delta = 3.31$ ppm). The term o.s. signifies overlapped signals. ¹³C NMR spectra were recorded in CDCl₃ or CD₃OD solution on the same apparatus at 75.3 MHz, and chemical shifts are reported with respect to solvent signals (CDCl₃: $\delta_{\text{C}} = 77.00$ ppm; CD₃OD: $\delta_{\text{C}} = 49.00$ ppm). ¹³C NMR multiplicities were determined by DEPT spectra. ¹H–¹H COSY, HSQC, HMBC and NOESY experiments were carried out with the same apparatus. Mass spectra (ESI) were recorded with a Shimadzu GC–MS–QP2010 Plus spectrometer and exact mass measurements were determined with a Micromass Autospec mass spectrometer at 70 eV. Optical rotations were recorded with a Jasco P-1010 polarimeter. Elemental analysis was carried out with a Perkin–Elmer 240 apparatus. Merck silica gel (70–230 mesh), deactivated with 15% H₂O, was used for column chromatography.

Isolation of (6*R*,7*S*,8*S*,1''*R*)-8-[(1',2''-Dihydroxyethyl)acroyloyl]-15-hydroxygermacra-1(10),4,11(13)-trien-6,12-olide(cnicin) (1**):** Cnicin (**1**) was isolated from *Centaurea napifolia* collected in June 2007 at Lascari, 60 km east of Palermo, Italy, and the purification procedures have been previously reported.^[22]

(6*R*,7*S*,8*S*)-6,15-Dihydroxy-germacra-1(10),4,11(13)-trien-8,12-olide (artemisiifolin) (2**):** Cnicin (5 g, 13.2 mmol) was dissolved in aqueous 10% KOH (500 mL) and stirred for 4 h. The reaction mixture was quenched with 10% H₂SO₄ and then extracted with EtOAc a few times. The combined organic extracts were dried with Na₂SO₄ (anhydrous) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel; petroleum ether/ethyl acetate, 3:7) to give artemisiifolin (2.7 g, 77%). The physical and spectroscopic data were identical to those reported in the literature.^[19,20]

(4*R*,5*S*,6*S*,7*S*,8*S*)-4,5-Epoxy-6,15-dihydroxygermacra-1(10),11(13)-dien-8,12-olide (3**) and (4*S*,5*R*,6*S*,7*S*,8*S*)-4,5-Epoxy-6,15-dihydroxygermacra-1(10),11(13)-dien-8,12-olide (**4**):** *m*-CIPBA (870 mg, 5.7 mmol) was added to a solution of artemisiifolin (**2**; 1 g, 3.8 mmol) in THF (50 mL) and pyridine (3.3 mL). The reaction mixture was stirred at room temperature for 24 h, quenched with a saturated aqueous solution of NH₄Cl and extracted with EtOAc. The organic layer was dried with anhydrous Na₂SO₄. After removal of the solvent, the residue was purified by column chromatography (silica gel; petroleum ether/EtOAc, 1:1) to give compounds **3** (560 mg, 53%) and **4** (160 mg, 15%). Data for **3**: Colourless oil. $[\alpha]_{\text{D}}^{24} = +98.4$ ($c = 0.43$, CHCl₃). IR (film): $\tilde{\nu} = 3421, 2928, 2862, 1757, 1277, 1144, 1013, 734$ cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 5.38$ (m, 1 H, 1-H), 2.38 (o.s., 2 H, 2-H^A and 2-H^B), 2.52 (ddd, $J = 12.9, 3.6, 3.6$ Hz, 1 H, 3-H^A), 1.12 (ddd, $J = 12.9, 9.9, 9.9$ Hz, 1 H, 3-H^B), 2.77 (o.s., 1 H, 5-H), 4.12 (dd, $J = 10.8, 3.3$ Hz, 1 H, 6-H), 2.77 (o.s., 1 H, 7-H), 4.42 (m, 1 H, 8-H), 2.77 (o.s., 1 H, 9-H^A), 1.99 (m, 1 H, 9-H^B), 6.44 (d, $J = 2.0$ Hz, 1 H, 13-H^A), 6.13 (d, $J = 1.2$ Hz, 1 H, 13-H^B), 1.76 (s, 3 H, 14-H₃), 4.14 (d, $J = 12.6$ Hz, 1 H, 15-H^A), 3.87 (d, $J = 12.6$ Hz, 1 H, 15-H^B) ppm. ¹³C

Table 1. ^{13}C NMR spectroscopic data for compounds **3–9** and **11–17** in CDCl_3 .

	3	4	5	6	7	8	9	11	12	13	14	15	16	17
C-1	127.3 (d)	126.9 (d)	88.5 (s)	126.3 (s)	43.0 (d)	127.8 (d)	126.8 (d)	61.7 (d)	65.4 (d)	67.2 (d)	70.8 (d)	78.4 (d)	77.7 (d)	77.9 (d)
C-2	22.9 (t)	22.5 (t)	33.8 (t)	29.7 (t)	36.5 (t)	23.0 (t)	22.6 (t)	23.6 (t)	23.1 (t)	27.9 (t)	29.7 (t)	28.1 (t)	25.6 (t)	27.4 (t)
C-3	32.5 (t)	31.5 (t)	33.2 (t)	34.1 (t)	26.7 (t)	31.7 (t)	30.9 (t)	29.5 (t)	32.0 (t)	32.3 (t)	41.8 (t)	33.9 (t)	35.0 (t)	34.8 (t)
C-4	64.7 (s)	69.9 (s)	83.0 (s)	83.5 (s)	83.0 (s)	62.3 (s)	62.8 (s)	139.3 (s)	137.6 (s)	135.9 (s)	136.7 (s)	136.2 (s)	72.0 (s)	73.5 (s)
C-5	65.6 (d)	66.6 (d)	57.9 (d)	58.8 (d)	67.0 (d)	60.5 (d)	66.0 (d)	128.7 (d)	129.1 (d)	130.3 (d)	130.6 (d)	131.7 (d)	52.0 (d)	56.1 (d)
C-6	68.3 (d)	69.7 (d)	70.2 (d)	70.8 (d)	73.2 (d)	68.5 (d)	70.4 (d)	72.3 (d)	71.1 (d)	62.2 (d)	71.9 (d)	69.6 (d)	69.5 (d)	69.4 (d)
C-7	46.8 (d)	48.8 (d)	51.6 (d)	54.9 (d)	48.8 (d)	44.5 (d)	47.5 (d)	46.4 (d)	51.4 (d)	49.2 (d)	48.2 (d)	51.1 (d)	53.8 (d)	54.1 (d)
C-8	77.2 (d)	80.7 (d)	79.2 (d)	77.0 (d)	76.8 (d)	77.2 (d)	80.3 (d)	77.3 (d)	76.5 (d)	74.0 (d)	78.3 (d)	76.4 (d)	76.6 (d)	75.9 (d)
C-9	43.4 (t)	44.3 (t)	36.6 (t)	39.9 (t)	40.3 (t)	43.5 (t)	44.5 (t)	44.2 (t)	46.6 (t)	46.6 (t)	31.4 (t)	125.8 (d)	41.5 (t)	43.4 (t)
C-10	130.0 (s)	130.5 (s)	33.5 (d)	131.6 (s)	144.7 (s)	130.2 (s)	130.3 (s)	56.7 (s)	57.9 (s)	142.8 (s)	146.6 (s)	149.0 (s)	42.1 (s)	41.4 (s)
C-11	134.3 (s)	133.8 (s)	137.7 (s)	137.6 (s)	136.6 (s)	133.8 (s)	133.3 (s)	135.0 (s)	134.8 (s)	136.7 (s)	135.6 (s)	136.7 (s)	136.7 (s)	136.3 (s)
C-12	170.0 (s)	170.0 (s)	170.2 (s)	170.1 (s)	170.2 (s)	168.7 (s)	168.3 (s)	168.4 (s)	168.6 (s)	169.1 (s)	169.0 (s)	169.2 (s)	169.7 (s)	169.5 (s)
C-13	128.2 (t)	128.9 (t)	124.1 (t)	124.3 (t)	126.6 (t)	127.7 (t)	127.5 (t)	126.4 (t)	126.6 (t)	122.9 (t)	125.5 (t)	121.8 (t)	118.9 (t)	119.3 (t)
C-14	19.0 (q)	19.0 (q)	20.5 (q)	23.6 (q)	112.4 (t)	19.6 (q)	18.7 (q)	21.4 (q)	17.4 (q)	16.7 (q)	115.8 (t)	12.0 (q)	15.4 (q)	16.8 (q)
C-15	59.6 (t)	61.4 (t)	73.0 (t)	64.2 (t)	66.1 (t)	61.6 (t)	62.6 (t)	62.7 (t)	61.6 (t)	61.7 (t)	62.2 (t)	60.6 (t)	71.4 (t)	64.7 (t)
Ac						170.7 (s)	170.2 (s)	169.5 (s)	170.5 (s)	170.8 (s)	169.6 (s)	170.6 (s)	172.2 (s)	170.9 (s)
Ac						168.7 (s)	168.9 (s)	170.5 (s)	169.3 (s)	169.8 (s)	170.6 (s)	169.6 (s)	171.0 (s)	170.4 (s)
						20.8 (q)	20.5 (q)	20.8 (q)	20.9 (q)	20.9 (q)	20.8 (q)	20.9 (q)	21.6 (q)	21.5 (q)
						20.5 (q)	20.4 (q)	20.8 (q)	20.9 (q)	20.9 (q)	20.8 (q)	20.9 (q)	20.9 (q)	20.8 (q)

NMR (75.3 MHz, CDCl_3): see Table 1. HRMS (ESI): calcd. for $\text{C}_{15}\text{H}_{20}\text{O}_5\text{Na}$ 303.12084; found 303.12078. Data for **4**: Colourless oil. $[\alpha]_D^{24} = +21.1$ ($c = 1.26$, CHCl_3). IR (film): $\tilde{\nu} = 3420, 2928, 2866, 1747, 1653, 1356, 1244, 1149, 1034, 918, 816\text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 5.36$ (m, 1 H, 1-H), 2.33 (o.s., 2 H, 2-H^A and 2-H^B), 2.33 (o.s., 1 H, 3-H^A), 1.15 (m, 1 H, 3-H^B), 2.76 (d, $J = 9.1$ Hz, 1 H, 5-H), 3.76 (dd, $J = 9.1, 9.1$ Hz, 1 H, 6-H), 2.91 (m, 1 H, 7-H), 3.97 (m, 1 H, 8-H), 2.70 (br. d, $J = 13.1$ Hz, 1 H, 9-H^A), 2.17 (dd, $J = 13.1, 11.3$ Hz, 1 H, 9-H^B), 6.41 (d, $J = 1.8$ Hz, 1 H, 13-H^A), 6.18 (br. s, 1 H, 13-H^B), 1.77 (s, 3 H, 14-H₃), 3.78 (d, $J = 12.3$ Hz, 1 H, 15-H^A), 3.57 (d, $J = 12.3$ Hz, 1 H, 15-H^B) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. HRMS (ESI): calcd. for $\text{C}_{15}\text{H}_{20}\text{O}_5\text{Na}$ 303.12084; found 303.12125.

(1S,4R,5S,6R,7S,8S,10S)-1,15-Epoxy-4,6-dihydroxyguaian-11(13)-en-8,12-olide (5), (4R,5S,6R,7S,8S)-4,6,15-Trihydroxyguaian-1(10),11(13)-dien-8,12-olide (6) and (1R,4R,5S,6R,7S,8S)-4,6,15-Trihydroxyguaian-10(14),11(13)-dien-8,12-olide (7): To a solution of **4** (50 mg, 0.18 mmol) dissolved in CHCl_3 (4 mL) was added a solution of CHCl_3 saturated with gaseous HCl (1 mL), at room temperature. After 4 d the solvent was evaporated and the mixture was subjected to column chromatography (silica gel; petroleum ether/EtOAc, 3:7) to give, in order of increasing polarity, compounds **5** (5 mg, 10%), **6** (10 mg, 20%) and **7** (12 mg, 25%). Data for **5**: Colourless oil. $[\alpha]_D^{24} = -29.3$ ($c = 0.08$, CHCl_3). IR (film): $\tilde{\nu} = 3397, 2925, 1752, 1636, 1341, 1278, 1018, 966\text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 1.89$ (m, 1 H, 2-H^A), 1.86 (m, 1 H, 2-H^B), 2.01 (m, 1 H, 3-H^A), 1.72 (m, 1 H, 3-H^B), 1.88 (d, $J = 8.7$ Hz, 1 H, 5-H), 4.18 (dd, $J = 10.2, 8.7$ Hz, 1 H, 6-H), 2.97 (dddd, $J = 10.2, 10.2, 3.0, 3.0$ Hz, 1 H, 7-H), 4.27 (ddd, $J = 10.2, 10.2, 4.2$ Hz, 1 H, 8-H), 2.38 (ddd, $J = 13.8, 6.3, 4.2$ Hz, 1 H, 9-H^A), 1.66 (ddd, $J = 13.8, 10.2, 6.6$ Hz, 1 H, 9-H^B), 2.12 (m, 1 H, 10-H), 6.29 (d, $J = 3.0$ Hz, 1 H, 13-H^A), 6.28 (d, $J = 3.0$ Hz, 1 H, 13-H^B), 1.15 (d, $J = 7.4$ Hz, 3 H, 14-H₃), 3.87 (dd, $J = 7.2, 3.3$ Hz, 1 H, 15-H^A), 3.70 (d, $J = 7.2$ Hz, 1 H, 15-H^B) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 319 (14) $[\text{M} + \text{K}]^+$, 303 (100) $[\text{M} + \text{Na}]^+$, 281 (4.5) $[\text{M} + \text{H}]^+$. $\text{C}_{15}\text{H}_{20}\text{O}_5$ (280.1311): calcd. C 64.27, H 7.19; found C 64.30, H 7.17. Data for **6**: Colourless oil. $[\alpha]_D^{24} = +98.4$ ($c = 0.43$, CHCl_3). IR (film): $\tilde{\nu} = 3321, 2910, 1758, 1635, 1260, 1055\text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 2.49$ (o.s., 1 H, 2-H^A), 2.09 (o.s., 1 H, 2-H^B), 2.09 (o.s., 1 H, 3-H^A), 1.56 (m, 1 H, 3-H^B), 2.73 (br. d, $J = 9.9$ Hz, 1 H, 5-H), 3.89 (dd, $J = 9.9, 9.9$ Hz,

1 H, 6-H), 2.83 (dddd, $J = 9.9, 9.9, 2.9, 2.9$ Hz, 1 H, 7-H), 3.83 (ddd, $J = 10.8, 9.9, 3.0$ Hz, 1 H, 8-H), 2.61 (dd, $J = 15.0, 3.0$ Hz, 1 H, 9-H^A), 2.49 (o.s., 1 H, 9-H^B), 6.33 (d, $J = 2.9$ Hz, 1 H, 13-H^A), 6.28 (d, $J = 2.9$ Hz, 1 H, 13-H^B), 1.75 (br. s, 3 H, 14-H₃), 3.93 (d, $J = 11.1$ Hz, 1 H, 15-H^A), 3.63 (d, $J = 11.1$ Hz, 1 H, 15-H^B) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 319 (24) $[\text{M} + \text{K}]^+$, 303 (100) $[\text{M} + \text{Na}]^+$. $\text{C}_{15}\text{H}_{20}\text{O}_5$ (280.1311): calcd. C 64.27, H 7.19; found C 64.24, H 7.21. Data for **7**: Amorphous solid. $[\alpha]_D^{24} = -48.2$ ($c = 0.19$, CHCl_3). IR (film): $\tilde{\nu} = 3318, 2921, 1756, 1640, 1265, 1041, 736\text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 2.17$ (m, 1 H, 1-H), 2.05 (m, 1 H, 2-H^A), 1.68 (m, 1 H, 2-H^B), 1.81 (m, 1 H, 3-H^A), 1.65 (m, 1 H, 3-H^B), 1.86 (dd, $J = 11.0$ Hz, 9.9, 1 H, 5-H), 3.98 (dd, $J = 9.9, 9.9$ Hz, 1 H, 6-H), 2.80 (dddd, $J = 9.9, 9.3, 3.1, 3.1$ Hz, 1 H, 7-H), 4.38 (ddd, $J = 11.1, 9.3, 6.3$ Hz, 1 H, 8-H), 3.16 (br. dd, $J = 15.3, 6.3$ Hz, 1 H, 9-H^A), 2.53 (dd, $J = 15.3, 11.1$ Hz, 1 H, 9-H^B), 6.37 (m, 2 H, 13-H^A and 13-H^B), 5.18 (d, $J = 2.1$ Hz, 14_A-H), 5.03 (d, $J = 1.5$ Hz, 14_B-H), 3.78 (d, $J = 11.6$ Hz, 1 H, 15-H^A), 3.63 (d, $J = 11.6$ Hz, 1 H, 15-H^B) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 319 (13) $[\text{M} + \text{K}]^+$, 303 (100) $[\text{M} + \text{Na}]^+$. $\text{C}_{15}\text{H}_{20}\text{O}_5$ (280.1311): calcd. C 64.27, H 7.19; found C 64.24, H 7.16.

(4R,5S,6S,7S,8S)-4,5-Epoxy-6,15-diacetoxygermacra-1(10),11(13)-dien-8,12-olide (8): To a stirred solution of compound **3** (100 mg, 0.36 mmol) in acetic anhydride (2 mL) was added, at room temperature, pyridine (2 mL). After 2 h, the reaction was concentrated and subjected to column chromatography (silica gel; petroleum ether/EtOAc, 7:3) to give compound **8** in quantitative yield. Colourless oil. $[\alpha]_D^{24} = +16.7$ ($c = 0.53$, CHCl_3). IR (film): $\tilde{\nu} = 2919, 2853, 1737, 1749, 1456, 1398, 1375, 1251, 1152, 1014, 968\text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 5.32$ (m, 1 H, 1-H), 2.33 (o.s., 2 H, 2-H^A and 2-H^B), 2.37 (m, 1 H, 3-H^A), 1.00 (m, 1 H, 3-H^B), 2.78 (o.s., 1 H, 5-H), 5.24 (dd, $J = 11.4, 3.0$ Hz, 1 H, 6-H), 2.80 (o.s., 1 H, 7-H), 4.52 (m, 1 H, 8-H), 2.78 (o.s., 1 H, 9-H^A), 1.94 (m, 1 H, 9-H^B), 6.26 (d, $J = 1.8$ Hz, 1 H, 13-H^A), 5.76 (d, $J = 1.2$ Hz, 1 H, 13-H^B), 1.69 (s, 3 H, 14-H₃), 4.30 (d, $J = 12.3$ Hz, 1 H, 15_A-H), 4.25 (d, $J = 12.3$ Hz, 1 H, 15_B-H), 2.04 (s, 3 H, Ac-H₃), 2.00 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 403 (19) $[\text{M} + \text{K}]^+$, 387 (100) $[\text{M} + \text{Na}]^+$. $\text{C}_{19}\text{H}_{24}\text{O}_7$ (364.1522): calcd. C 62.63, H 6.64; found C 62.67, H 6.68.

(4S,5R,6S,7S,8S)-4,5-Epoxy-6,15-diacetoxygermacra-1(10),11(13)-dien-8,12-olide (9): Acetic anhydride (2 mL) was added to a solution of compound **4** (100 mg, 0.36 mmol) in pyridine (2 mL) at room temperature. After 2 h, the reaction was concentrated and subjected to column chromatography (silica gel; petroleum ether/EtOAc, 7:3) to give compound **9** in quantitative yield. Colourless oil. $[\alpha]_D^{24} = +55.5$ ($c = 0.18$, CHCl_3). IR (film): $\tilde{\nu} = 3502, 2926, 2868, 1732, 1653, 1439, 1372, 1339, 1224, 1122, 984, 816 \text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 5.35$ (br. dd, $J = 7.2, 7.2 \text{ Hz}$, 1 H, 1-H), 2.32 (o.s., 2 H, 2-H^A and 2-H^B), 2.32 (o.s., 1 H, 3-H^A), 1.24 (m, 1 H, 3-H^B), 2.83 (d, $J = 9.3 \text{ Hz}$, 1 H, 5-H), 4.94 (dd, $J = 9.3, 9.3 \text{ Hz}$, 1 H, 6-H), 3.12 (dddd, $J = 9.3, 4.2, 2.4, 2.4 \text{ Hz}$, 1 H, 7-H), 4.08 (ddd, $J = 10.5, 4.2, 1.8 \text{ Hz}$, 1 H, 8-H), 2.70 (br. d, $J = 10.5 \text{ Hz}$, 1 H, 9-H^A), 2.23 (br. dd, $J = 10.5, 10.5 \text{ Hz}$, 1 H, 9-H^B), 6.39 (d, $J = 2.4 \text{ Hz}$, 1 H, 13-H^A), 5.83 (d, $J = 2.4 \text{ Hz}$, 1 H, 13-H^B), 1.83 (s, 3 H, 14-H₃), 4.54 (d, $J = 12.0 \text{ Hz}$, 1 H, 15-H^A), 3.84 (d, $J = 12.0 \text{ Hz}$, 1 H, 15-H^B), 2.11 (s, 3 H, Ac-H₃), 2.07 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 403 (14) $[\text{M} + \text{K}]^+$, 387 (100) $[\text{M} + \text{Na}]^+$, 365 (2) $[\text{M} + \text{H}]^+$. $\text{C}_{19}\text{H}_{24}\text{O}_7$ (364.1522): calcd. C 62.63, H 6.64; found C 62.59, H 6.62.

(6R,7S,8S)-6,15-Diacetoxygermacra-1(10),4,11(13)-trien-8,12-olide (dicetylartemisiifolin) (10): Artemisiifolin (1.6 g, 6.1 mmol) was dissolved in pyridine (25 mL) and acetic anhydride (30 mL) was added at room temperature. After 1 h, the mixture was concentrated and subjected to column chromatography (silica gel; petroleum ether/EtOAc, 6:4) to give **10** in quantitative yield.^[25] Colourless oil. $[\alpha]_D^{24} = +147.6$ ($c = 1.72$, CHCl_3). HRMS (ESI): calcd. for $\text{C}_{19}\text{H}_{24}\text{O}_6\text{Na}$ 371.14706; found 371.14686.

(1S,6R,7S,8S,10S)-1,10-Epoxy-6,15-diacetoxygermacra-4,11(13)-dien-8,12-olide (11) and (1R,6R,7S,8S,10R)-1,10-Epoxy-6,15-diacetoxygermacra-4,11(13)-dien-8,12-olide (12): To a solution of dicetylartemisiifolin (**10**; 1.6 g, 4.6 mmol) dissolved in THF (35 mL) was added *m*-CIPBA (870 mg, 5.0 mmol) and pyridine (3 mL). The mixture was stirred for 48 h at room temperature and then washed in turn with saturated aqueous NaHCO_3 and NH_4Cl . The organic layer was separated, dried with Na_2SO_4 anhydrous and concentrated. The residue was purified by TLC plates eluted a few times with petroleum ether/diethyl ether (1:4) to give compounds **11** (420 mg, 25%) and **12** (490 mg, 29%). Data for **11**: Colourless oil. $[\alpha]_D^{24} = +128.2$ ($c = 0.63$, CHCl_3). IR (film): $\tilde{\nu} = 2933, 1764, 1738, 1373, 1234, 1147, 1026 \text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 2.69$ (dd, $J = 10.2, 1.5 \text{ Hz}$, 1 H, 1-H), 2.14 (m, 1 H, 2-H^A), 1.65 (m, 1 H, 2-H^B), 2.53 (ddd, $J = 15.4, 7.0, 3.0 \text{ Hz}$, 1 H, 3-H^A), 2.34 (ddd, $J = 15.4, 9.3, 7.5 \text{ Hz}$, 1 H, 3-H^B), 5.32 (o.s., 1 H, 5-H), 5.33 (o.s., 1 H, 6-H), 3.19 (m, 1 H, 7-H), 4.32 (ddd, $J = 7.8, 7.8, 4.8 \text{ Hz}$, 1 H, 8-H), 2.10 (o.s., 2 H, 9-H^A and 9-H^B), 6.38 (d, $J = 2.6 \text{ Hz}$, 1 H, 13-H^A), 5.91 (d, $J = 2.2 \text{ Hz}$, 1 H, 13-H^B), 1.47 (s, 3 H, 14-H₃), 4.96 (d, $J = 12.6 \text{ Hz}$, 1 H, 15-H^A), 4.59 (d, $J = 12.6 \text{ Hz}$, 1 H, 15-H^B), 2.10 (s, 6 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. HRMS (ESI): calcd. for $\text{C}_{19}\text{H}_{24}\text{O}_7\text{Na}$ 387.14197; found 387.14266. Data for **12**: Colourless oil. $[\alpha]_D^{24} = +12.0$ ($c = 0.42$, CHCl_3). IR (film): $\tilde{\nu} = 2935, 1762, 1740, 1375, 1275, 1149, 1026, 960, 735 \text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 2.71$ (br. d, $J = 9.5 \text{ Hz}$, 1 H, 1-H), 2.10 (m, 1 H, 2-H^A), 1.48 (m, 1 H, 2-H^B), 2.46 (br. dd, $J = 13.0, 3.4 \text{ Hz}$, 1 H, 3-H^A), 2.34 (ddd, $J = 13.0, 6.2, 6.0 \text{ Hz}$, 1 H, 3-H^B), 5.31 (d, $J = 9.6 \text{ Hz}$, 1 H, 5-H), 5.42 (dd, $J = 9.8, 9.8 \text{ Hz}$, 1 H, 6-H), 3.07 (dddd, $J = 9.8, 7.8, 2.8, 1.8 \text{ Hz}$, 1 H, 7-H), 4.15 (br. dd, $J = 9.0, 7.8 \text{ Hz}$, 1 H, 8-H), 2.70 (br. d, $J = 14.0 \text{ Hz}$, 1 H, 9-H^A), 1.57 (dd, $J = 14.0, 9.0 \text{ Hz}$, 1 H, 9-H^B), 6.37 (d, $J = 2.8 \text{ Hz}$, 1 H, 13-H^A), 5.92 (d, $J = 1.6 \text{ Hz}$, 1 H, 13-H^B), 1.31 (s, 3 H, 14-H₃), 4.79 (s, 2 H, 15-H^A and 15-H^B), 2.10 (s, 3 H, Ac-H₃), 2.09 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see

Table 1. HRMS (ESI): calcd. for $\text{C}_{19}\text{H}_{24}\text{O}_7\text{Na}$ 387.14197; found 387.14254.

(1S,6R,7S,8S)-1-Hydroxy-6,15-diacetoxygermacra-4,9,11(13)-trien-8,12-olide (13) and (1S,6R,7S,8S)-1-Hydroxy-6,15-diacetoxygermacra-4,10(14),11(13)-trien-8,12-olide (14): To a solution of **11** (190 mg, 0.52 mmol) dissolved in CHCl_3 (15 mL) was added a solution of CHCl_3 saturated with gaseous HCl (4 mL), at room temperature. After 24 h the solvent was evaporated and the mixture was subjected to column chromatography (silica gel; petroleum ether/EtOAc, 7:3) to give compounds **13** (95 mg, 50%) and **14** (20 mg, 10%). Data for **13**: Amorphous solid. $[\alpha]_D^{24} = -6.65$ ($c = 0.41$, CHCl_3). IR (film): $\tilde{\nu} = 3447, 2949, 2866, 1740, 1236, 1138, 1020, 735 \text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 4.49$ (dd, $J = 10.1, 4.8 \text{ Hz}$, 1 H, 1-H), 1.95 (o.s., 1 H, 2-H^A), 1.75 (m, 1 H, 2-H^B), 2.52 (m, 1 H, 3-H^A), 1.95 (o.s., 1 H, 3-H^B), 5.10 (d, $J = 9.6 \text{ Hz}$, 1 H, 5-H), 5.47 (dd, $J = 9.6, 9.6 \text{ Hz}$, 1 H, 6-H), 3.06 (m, 1 H, 7-H), 4.77 (dd, $J = 9.6, 9.6 \text{ Hz}$, 1 H, 8-H), 5.32 (d, $J = 9.6 \text{ Hz}$, 1 H, 9-H), 6.29 (d, $J = 3.0 \text{ Hz}$, 1 H, 13-H^A), 5.75 (d, $J = 3.0 \text{ Hz}$, 1 H, 13-H^B), 1.84 (s, 3 H, 14-H₃), 4.99 (d, $J = 12.8 \text{ Hz}$, 1 H, 15-H^A), 4.81 (d, $J = 12.8 \text{ Hz}$, 1 H, 15-H^B), 2.12 (s, 3 H, Ac-H₃), 2.09 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 751 (100) $[\text{2M} + \text{Na}]^+$, 403 (16) $[\text{M} + \text{K}]^+$, 387 (60) $[\text{M} + \text{Na}]^+$. $\text{C}_{19}\text{H}_{24}\text{O}_7$ (364.1522): calcd. C 62.63, H 6.64; found C 62.67, H 6.69. Data for **14**: Colourless oil. $[\alpha]_D^{24} = +84.6$ ($c = 0.09$, CHCl_3). IR (film): $\tilde{\nu} = 2924, 1736, 1496, 1234, 1150, 1026 \text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 3.90$ (dd, $J = 7.6, 7.6 \text{ Hz}$, 1 H, 1-H), 2.10 (o.s., 1 H, 2-H^A), 1.26 (m, 1 H, 2-H^B), 2.40 (m, 1 H, 3-H^A), 2.27 (o.s., 1 H, 3-H^B), 5.19 (o.s., 1 H, 5-H), 5.24 (o.s., 1 H, 6-H), 3.04 (m, 1 H, 7-H), 4.16 (m, 1 H, 8-H), 2.27 (o.s., 1 H, 9-H^A), 2.10 (o.s., 1 H, 9-H^B), 6.38 (d, $J = 2.9 \text{ Hz}$, 1 H, 13-H^A), 5.89 (d, $J = 2.5 \text{ Hz}$, 1 H, 13-H^B), 5.21 (o.s., 2 H, 14-H^A and 14-H^B), 4.83 (d, $J = 13.0 \text{ Hz}$, 1 H, 15-H^A), 4.68 (d, $J = 13.0 \text{ Hz}$, 1 H, 15-H^B), 2.11 (s, 3 H, Ac-H₃), 2.09 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 751 (100) $[\text{2M} + \text{Na}]^+$, 403 (40) $[\text{M} + \text{K}]^+$, 387 (68) $[\text{M} + \text{Na}]^+$. $\text{C}_{19}\text{H}_{24}\text{O}_7$ (364.1522): calcd. C 62.63, H 6.64; found C 62.61, H 6.62.

(1R,6R,7S,8S)-1-Hydroxy-6,15-diacetoxygermacra-4,9,11(13)-trien-8,12-olide (15), (1R,4S,5S,6R,7S,8S,10R)-1,4-Dihydroxy-6,15-diacetoxyeudesma-11(13)-en-8,12-olide (16) and (1R,4R,5S,6R,7S,8S,10R)-1,4-Dihydroxy-6,15-diacetoxyeudesma-11(13)-en-8,12-olide (17): A solution of CHCl_3 saturated with gaseous HCl (4 mL) was added to a solution of compound **12** (220 mg, 0.60 mmol) in CHCl_3 (15 mL) whilst stirring. After 24 h the solvent was evaporated and the mixture was subjected to column chromatography (silica gel; petroleum ether/EtOAc, 2:3) to give compounds **15** (11 mg, 0.03 mmol, 5%), **16** (31 mg, 0.08 mmol, 13%) and **17** (100 mg, 0.26 mmol, 43%). Data for **15**: Colourless oil. $[\alpha]_D^{24} = +17.0$ ($c = 0.12$, CHCl_3). IR (film): $\tilde{\nu} = 3435, 2924, 2854, 1718, 1375, 1256, 1142, 1036 \text{ cm}^{-1}$. ^1H NMR (300 MHz, CDCl_3): $\delta = 4.38$ (dd, $J = 10.0, 5.0 \text{ Hz}$, 1 H, 1-H), 1.77 (o.s., 2 H, 2-H^A and 2-H^B), 2.52 (m, 1 H, 3-H^A), 2.19 (m, 1 H, 3-H^B), 5.03 (br. d, $J = 10.3 \text{ Hz}$, 1 H, 5-H), 5.51 (dd, $J = 10.3, 10.3 \text{ Hz}$, 1 H, 6-H), 3.02 (dddd, $J = 10.3, 10.0, 3.4, 3.0 \text{ Hz}$, 1 H, 7-H), 4.60 (dd, $J = 10.0, 9.5 \text{ Hz}$, 1 H, 8-H), 5.36 (d, $J = 9.5 \text{ Hz}$, 1 H, 9-H), 6.28 (d, $J = 3.4 \text{ Hz}$, 1 H, 13-H^A), 5.72 (d, $J = 3.0 \text{ Hz}$, 1 H, 13-H^B), 1.78 (s, 3 H, 14-H₃), 4.83 (d, $J = 13.2 \text{ Hz}$, 1 H, 15-H^A), 4.69 (d, $J = 13.2 \text{ Hz}$, 1 H, 15-H^B), 2.11 (s, 3 H, Ac-H₃), 2.10 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 403 (8) $[\text{M} + \text{K}]^+$, 387 (100) $[\text{M} + \text{Na}]^+$. $\text{C}_{19}\text{H}_{24}\text{O}_7$ (364.1522): calcd. C 62.63, H 6.64; found C 62.70, H 6.63. Data for **16**: Amorphous solid. $[\alpha]_D^{24} = -4.9$ ($c = 0.22$, CHCl_3). IR (film): $\tilde{\nu} = 3398, 2923, 2854, 1743, 1716, 1653, 1458, 1259, 1064 \text{ cm}^{-1}$. ^1H NMR

(300 MHz, CDCl_3): δ = 3.43 (dd, J = 11.5, 4.0 Hz, 1 H, 1-H), 1.98 (m, 1 H, 2-H^A), 1.65 (m, 1 H, 2-H^B), 1.87 (m, 1 H, 3-H^A), 1.63 (o.s., 1 H, 3-H^B), 1.76 (d, J = 10.4 Hz, 1 H, 5-H), 5.64 (dd, J = 10.4, 10.4 Hz, 1 H, 6-H), 2.80 (dddd, J = 11.9, 10.4, 2.9, 2.9 Hz, 1 H, 7-H), 4.11 (ddd, J = 11.9, 11.9, 3.9 Hz, 1 H, 8-H), 2.58 (dd, J = 11.9, 3.9 Hz, 1 H, 9-H^A), 1.45 (dd, J = 11.9, 11.9 Hz, 1 H, 9-H^B), 6.12 (d, J = 2.9 Hz, 1 H, 13-H^A), 5.35 (d, J = 2.9 Hz, 1 H, 13-H^B), 1.23 (s, 3 H, 14-H₃), 3.85 (s, 2 H, 15-H^A and 15-H^B), 2.20 (s, 3 H, Ac-H₃), 2.11 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 421 (55) [$\text{M} + \text{K}$]⁺, 405 (100) [$\text{M} + \text{Na}$]⁺. $\text{C}_{19}\text{H}_{26}\text{O}_8$ (382.1628): calcd. C 59.68, H 6.85; found C 59.72, H 6.81. Data for **17**: Amorphous solid. $[a]_D^{25}$ = -4.1 (c = 0.66, CHCl_3). IR (film): $\tilde{\nu}$ = 3445, 2945, 2875, 1732, 1670, 1369, 1251, 1138, 735 cm^{-1} . ^1H NMR (300 MHz, CDCl_3): δ = 3.53 (dd, J = 10.7, 5.0 Hz, 1 H, 1-H), 1.71 (m, 2 H, 2-H^A and 2-H^B), 1.97 (ddd, J = 13.4, 3.4, 3.4 Hz, 1 H, 3-H^A), 1.46 (o.s., 1 H, 3-H^B), 2.03 (d, J = 10.4 Hz, 1 H, 5-H), 5.65 (dd, J = 10.4, 10.4 Hz, 1 H, 6-H), 2.83 (dddd, J = 11.9, 10.4, 2.9, 2.9 Hz, 1 H, 7-H), 4.01 (ddd, J = 11.9, 11.9, 3.7 Hz, 1 H, 8-H), 2.59 (dd, J = 11.9, 3.7 Hz, 1 H, 9-H^A), 1.51 (dd, J = 11.9, 11.9 Hz, 1 H, 9-H^B), 6.13 (d, J = 2.9 Hz, 1 H, 13-H^A), 5.38 (d, J = 2.9 Hz, 1 H, 13-H^B), 1.06 (s, 3 H, 14-H₃), 4.38 (d, J = 11.9 Hz, 1 H, 15-H^A), 4.35 (d, J = 11.9 Hz, 1 H, 15-H^B), 2.16 (s, 3 H, Ac-H₃), 2.12 (s, 3 H, Ac-H₃) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 1. MS (ESI⁺): m/z (%) = 421 (56) [$\text{M} + \text{K}$]⁺, 405 (100) [$\text{M} + \text{Na}$]⁺, 383 (43) [$\text{M} + \text{H}$]⁺. $\text{C}_{19}\text{H}_{26}\text{O}_8$ (382.1628): calcd. C 59.68, H 6.85; found C 59.65, H 6.87.

(1S,6R,7S,8S,10S,11S)-1,10-Epoxy-6,15-dihydroxy-13-methoxygermacra-4-en-8,12-olide (18): To a solution of **11** (208 mg, 0.57 mmol) dissolved in CH_3OH (10 mL) was added NaHCO_3 (240 mg, 2.8 mmol). The mixture was stirred for 24 h at room temperature, quenched with water and extracted with EtOAc. The combined organic layers were dried with Na_2SO_4 and evaporated to give compound **18** (160 mg, 0.51 mmol, 90%). Colourless oil. $[a]_D^{25}$ = +86.5 (c = 0.27, CHCl_3). IR (film): $\tilde{\nu}$ = 3396, 2930, 2870, 1751, 1424, 1387, 1197, 1029, 756 cm^{-1} . ^1H NMR (300 MHz, CDCl_3): δ = 2.79 (dd, J = 10.5, 2.7 Hz, 1 H, 1-H), 2.14 (o.s., 1 H, 2-H^A), 1.53 (m, 1 H, 2-H^B), 2.35 (o.s., 1 H, 3-H^A), 2.35 (o.s., 1 H, 3-H^B), 5.36 (d, J = 8.1 Hz, 1 H, 5-H), 4.34 (dd, J = 9.9, 8.1, 10.4 Hz, 1 H, 6-H), 2.53 (ddd, J = 8.1, 5.7, 5.7 Hz, 1 H, 7-H), 4.32 (m, 1 H, 8-H), 2.16 (dd, J = 13.6, 11.1 Hz, 1 H, 9-H^A), 1.99 (dd, J = 13.6, 4.5 Hz, 1 H, 9-H^B), 2.89 (ddd, J = 5.7, 5.7, 3.7 Hz, 1 H, 11-H), 3.72 (dd, J = 9.0, 3.7 Hz, 1 H, 13-H^A), 3.67 (dd, J = 9.0, 5.7 Hz, 1 H, 13-H^B), 1.43 (s, 3 H, 14-H₃), 4.23 (d, J = 13.2 Hz, 1 H, 15-H^A), 4.14 (d, J

= 13.2 Hz, 1 H, 15-H^B), 3.42 (s, 3 H, $\text{OCH}_3\text{-H}_3$) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 2. MS (ESI⁺): m/z (%) = 351 (7) [$\text{M} + \text{K}$]⁺, 335 (100) [$\text{M} + \text{Na}$]⁺. $\text{C}_{16}\text{H}_{24}\text{O}_6$ (312.1573): calcd. C 61.52, H 7.74; found C 61.55, H 7.70.

(1S,6R,7S,8S,10S)-1,10-Epoxy-6,15-dihydroxygermacra-4,11(13)-dien-8,12-olide (19): To a solution of **18** (155 mg, 0.50 mmol) dissolved in dry THF (8 mL) was added $t\text{BuOK}$ (66 mg, 0.60 mmol). The mixture was stirred for 10 min at room temperature, then quenched with a saturated solution of NH_4Cl and extracted with EtOAc. The combined organic layers were dried with Na_2SO_4 and evaporated to give compound **19** (80 mg, 0.28 mmol, 56%). Data for **19**: Colourless oil. $[a]_D^{25}$ = +161.5 (c = 1.49, CH_3OH). IR (film): $\tilde{\nu}$ = 3415, 3340, 1761, 1458, 1280, 1159, 1034, 1017 cm^{-1} . ^1H NMR (300 MHz, CDCl_3): δ = 2.82 (dd, J = 10.2, 2.1 Hz, 1 H, 1-H), 2.15 (o.s., 1 H, 2-H^A), 1.53 (m, 1 H, 2-H^B), 2.34 (o.s., 1 H, 3-H^A), 2.34 (o.s., 1 H, 3-H^B), 5.47 (d, J = 7.8 Hz, 1 H, 5-H), 4.32 (o.s., J = 10.4 Hz, 1 H, 6-H), 3.12 (m, 1 H, 7-H), 4.35 (o.s., 1 H, 8-H), 2.07 (o.s., 1 H, 9-H^A), 2.07 (o.s., 1 H, 9-H^B), 6.37 (d, J = 2.4 Hz, 1 H, 13-H^A), 6.15 (br. s, 1 H, 13-H^B), 1.44 (s, 3 H, 14-H₃), 4.24 (s, 2 H, 15_A-H and 15_B-H) ppm. ^{13}C NMR (75.3 MHz, CDCl_3): see Table 2. MS (ESI⁺): m/z (%) = 319 (8) [$\text{M} + \text{K}$]⁺, 303 (100) [$\text{M} + \text{Na}$]⁺. $\text{C}_{15}\text{H}_{20}\text{O}_5$ (280.1311): calcd. C 64.27, H 7.19; found C 64.31, H 7.16.

(1S,4R,5R,6R,7S,8S,10S,15R)-1,15-Dihydroxy-6,15-epoxyeudesma-11(13)-en-8,12-olide (20), **(1S,5R,6R,7S,8S,10S)-1-Hydroxy-6,15-epoxyeudesma-4(15),11(13)-dien-8,12-olide (21)** and **(1S,6R,7S,8S)-1,6,15-Trihydroxygermacra-9,11(13)-dien-8,12-olide (22)**: A solution of CHCl_3 saturated with gaseous HCl (2 mL) was added to a solution of compound **19** (80 mg, 0.29 mmol) in CHCl_3 (8 mL), whilst stirring. After 24 h, the solvent was evaporated, and the mixture was subjected to column chromatography (silica gel; petroleum ether/EtOAc, 3:7) to give compounds **20** (11 mg, 0.04 mmol, 14%), **21** (5 mg, 0.02 mmol, 6%) and **22** (28 mg, 0.1 mmol, 34%), whereas **19** (15 mg, 0.05 mmol) was also recovered. Data for **20**: Colourless oil. $[a]_D^{25}$ = -10.13 (c = 0.25, CHCl_3). IR (film): $\tilde{\nu}$ = 3413, 2924, 1750, 1455, 1254, 1135, 1029, 737 cm^{-1} . ^1H NMR (300 MHz, CDCl_3): δ = 3.51 (dd, J = 9.4, 5.6 Hz, 1 H, 1-H), 1.82 (m, 1 H, 2-H^A), 1.62 (m, 1 H, 2-H^B), 1.95 (m, 1 H, 3-H^A), 1.27 (o.s., 1 H, 3-H^B), 2.26 (o.s., 1 H, 4-H), 2.05 (dd, J = 5.5, 2.2 Hz, 1 H, 5-H), 4.57 (dd, J = 7.2, 5.0 Hz, 1 H, 6-H), 3.38 (dddd, J = 12.6, 7.2, 3.5, 3.0 Hz, 1 H, 7-H), 4.09 (ddd, J = 12.6, 9.6, 8.4 Hz, 1 H, 8-H), 2.26 (dd, J = 13.4, 8.4 Hz, 1 H, 9-H^A), 1.83 (dd, J = 13.4, 9.6 Hz, 1 H,

Table 2. ^{13}C NMR spectroscopic data for compounds **18–26** in CDCl_3 .

	18	19	20	21	22 ^[a]	23	24 ^[b]	25	26 ^[b]
C-1	59.2 (d)	61.6 (d)	77.2 (d)	77.8 (d)	68.0 (d)	66.5 (d)	65.5 (d)	77.2 (d)	77.2 (d)
C-2	24.2 (t)	23.8 (t)	27.9 (t)	30.4 (t)	29.0 (t)	23.4 (t)	23.0 (t)	29.1 (t)	26.3 (t)
C-3	30.8 (t)	30.6 (t)	19.5 (t)	20.4 (t)	32.9 (t)	31.8 (t)	31.9 (t)	32.8 (t)	22.6 (t)
C-4	140.6 (s)	140.0 (s)	44.1 (d)	112.6 (s)	138.1 (s)	131.2 (s)	138.4 (s)	60.9 (s)	43.3 (d)
C-5	129.2 (d)	128.9 (d)	47.0 (d)	51.3 (d)	133.9 (d)	139.4 (d)	131.8 (d)	51.5 (d)	54.2 (d)
C-6	71.5 (d)	69.6 (d)	78.4 (d)	80.3 (d)	70.6 (d)	69.4 (d)	68.8 (d)	67.8 (d)	66.4 (d)
C-7	51.0 (d)	49.7 (d)	46.6 (d)	45.2 (d)	53.5 (d)	55.8 (d)	53.8 (d)	54.3 (d)	55.3 (d)
C-8	78.7 (d)	77.5 (d)	75.9 (d)	75.8 (d)	76.4 (d)	78.1 (d)	77.0 (d)	76.0 (d)	77.7 (d)
C-9	44.4 (t)	43.9 (t)	38.4 (t)	35.3 (t)	127.4 (d)	45.9 (t)	46.3 (t)	40.8 (t)	41.2 (t)
C-10	61.7 (s)	57.2 (s)	37.4 (s)	39.0 (s)	143.7 (s)	58.0 (s)	58.1 (s)	43.0 (s)	41.3 (s)
C-11	47.5 (d)	135.5 (s)	138.2 (s)	138.2 (s)	140.2 (s)	47.5 (d)	135.8 (s)	136.9 (s)	137.5 (s)
C-12	176.1 (s)	170.0 (s)	170.2 (s)	170.1 (s)	172.3 (s)	174.7 (s)	170.0 (s)	170.2 (s)	171.0 (s)
C-13	72.3 (t)	126.4 (t)	119.6 (t)	120.4 (t)	122.9 (t)	71.6 (t)	126.9 (t)	120.6 (t)	119.7 (t)
C-14	21.1 (q)	20.7 (q)	21.2 (q)	19.0 (q)	17.0 (q)	17.5 (q)	17.2 (q)	14.3 (q)	14.8 (q)
C-15	61.7 (t)	61.0 (t)	104.8 (d)	138.1 (d)	59.8 (t)	60.4 (t)	59.8 (t)	51.9 (t)	204.7 (d)
OCH_3	57.2 (q)					59.2 (q)			

[a] In MeOD. [b] In $\text{CDCl}_3/\text{MeOD}$, 9:1.

9-H^B), 6.21 (d, $J = 3.5$ Hz, 1 H, 13-H^A), 5.76 (d, $J = 3.0$ Hz, 1 H, 13-H^B), 1.27 (s, 3 H, 14-H₃), 5.43 (d, $J = 6.0$ Hz, 1 H, 15-H) ppm. ¹³C NMR (75.3 MHz, CDCl₃): see Table 2. MS (ESI⁺): m/z (%) = 303 (3) [M + Na]⁺, 285 (100) [M – H₂O + Na]⁺. C₁₅H₂₀O₅ (280.1311): calcd. C 64.27, H 7.19; found C 64.30, H 7.17. Data for **21**: Colourless oil. $[α]_D^{24} = -32.34$ ($c = 0.15$, CHCl₃). IR (film): $\tilde{\nu} = 3416, 2926, 1767, 1666, 1453, 1256, 1192, 1024, 737$ cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 3.52$ (o.s., 1 H, 1-H), 1.79 (m, 1 H, 2-H^A), 1.49 (m, 1 H, 2-H^B), 2.47 (m, 1 H, 3-H^A), 2.24 (m, 1 H, 3-H^B), 2.35 (br. d, $J = 9.2$ Hz, 1 H, 5-H), 4.50 (dd, $J = 9.2, 9.2$ Hz, 1 H, 6-H), 3.55 (o.s., 1 H, 7-H), 4.04 (ddd, $J = 12.3, 9.3, 8.8$ Hz, 1 H, 8-H), 2.20 (dd, $J = 13.4, 8.8$ Hz, 1 H, 9-H^A), 1.92 (dd, $J = 13.4, 9.3$ Hz, 1 H, 9-H^B), 6.24 (d, $J = 3.0$ Hz, 1 H, 13-H^A), 5.77 (d, $J = 3.0$ Hz, 1 H, 13-H^B), 1.12 (s, 3 H, 14-H₃), 6.18 (m, 1 H, 15-H) ppm. ¹³C NMR (75.3 MHz, CDCl₃): see Table 2. MS (ESI⁺): m/z (%) = 301 (57) [M + K]⁺, 285 (100) [M + Na]⁺. C₁₅H₁₈O₄ (262.1205): calcd. C 68.68, H 6.92; found C 68.70, H 6.89. Data for **22**: Colourless oil. $[α]_D^{24} = -49.85$ ($c = 0.50$, CH₃OH). IR (film): $\tilde{\nu} = 3420, 3333, 1763, 1275, 1165, 1040$ cm⁻¹. ¹H NMR (300 MHz, CD₃OD): $\delta = 4.52$ (dd, $J = 12.0, 5.1$ Hz, 1 H, 1-H), 2.02 (m, 1 H, 2-H^A) 1.74 (m, 1 H, 2-H^B), 2.58 (br. dd, $J = 12.6, 6.3$ Hz, 1 H, 3-H^A), 1.87 (o.s., 1 H, 3-H^B), 5.06 (d, $J = 10.8$ Hz, 1 H, 5-H), 4.57 (dd, $J = 10.8, 9.0$ Hz, 1 H, 6-H), 2.79 (dddd, $J = 9.0, 9.0, 3.3, 3.0$ Hz, 1 H, 7-H), 4.78 (dd, $J = 10.3, 9.0$ Hz, 1 H, 8-H), 5.31 (dd, $J = 10.3, 1.3$ Hz, 1 H, 9-H), 6.17 (o.s., 1 H, 13-H^A), 6.17 (o.s., 1 H, 13-H^B), 1.83 (br. s, 3 H, 14-H₃), 4.28 (d, $J = 12.6$ Hz, 1 H, 15-H^A), 4.15 (d, $J = 12.6$ Hz, 1 H, 15-H^B) ppm. ¹³C NMR (75.3 MHz, CD₃OD): see Table 2. MS (ESI⁺): m/z (%) = 303 (100) [M + Na]⁺. C₁₅H₂₀O₅ (280.1311): calcd. C 64.27, H 7.19; found C 64.24, H 7.23.

(1R,6R,7S,8S,10R,11S)-1,10-Epoxy-6,15-dihydroxy-13-methoxygermacra-4-en-8,12-olide (23): To a solution of **12** (110 mg, 0.30 mmol) dissolved in CH₃OH (40 mL) was added NaHCO₃ (125 mg, 1.48 mmol). The mixture was stirred at room temperature for 24 h, quenched with water and extracted with EtOAc. The combined organic layers were dried with Na₂SO₄ and evaporated to give compound **23** (90 mg, 0.29 mmol, 96%). Colourless oil. $[α]_D^{24} = -8.39$ ($c = 0.20$, CHCl₃). IR (film): $\tilde{\nu} = 3384, 2930, 1761, 1450, 1364, 1299, 1160, 935, 754$ cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 2.74$ (dd, $J = 11.1, 2.1$ Hz, 1 H, 1-H), 2.08 (m, 2 H, 2-H^A), 1.38 (m, 1 H, 2-H^B), 2.50 (ddd, $J = 12.9, 5.4, 2.7$ Hz, 1 H, 3-H^A), 2.28 (m, 1 H, 3-H^B), 5.31 (d, $J = 9.6$ Hz, 1 H, 5-H), 4.51 (dd, $J = 9.6, 9.6$ Hz, 1 H, 6-H), 2.38 (ddd, $J = 9.6, 9.6, 9.6$ Hz, 1 H, 7-H), 4.20 (br. dd, $J = 9.6, 9.6$ Hz, 1 H, 8-H), 2.67 (br. d, $J = 14.1$ Hz, 1 H, 9-H^A), 1.52 (dd, $J = 14.1, 9.6$ Hz, 1 H, 9-H^B), 2.92 (ddd, $J = 9.6, 6.6, 3.3$ Hz, 1 H, 11-H), 3.94 (dd, $J = 9.0, 3.3$ Hz, 1 H, 13-H^A), 3.59 (dd, $J = 9.0, 6.6$ Hz, 1 H, 13-H^B), 1.20 (s, 3 H, 14-H₃), 4.35 (d, $J = 12.9$ Hz, 1 H, 15-H^A), 4.01 (d, $J = 12.9$ Hz, 1 H, 15-H^B), 3.44 (s, 3 H, OCH₃-H₃) ppm. ¹³C NMR (75.3 MHz, CDCl₃): see Table 2. MS (ESI⁺): m/z (%) = 351 (30) [M + K]⁺, 335 (100) [M + Na]⁺. C₁₆H₂₄O₆ (312.1573): calcd. C 61.52, H 7.74; found C 61.48, H 7.75.

(1R,6R,7S,8S,10R)-1,10-Epoxy-6,15-dihydroxygermacra-4,11(13)-dien-8,12-olide (24): To a solution of **23** (72 mg, 0.23 mmol) dissolved in dry THF (6 mL) was added *t*BuOK (30 mg, 0.26 mmol). The mixture was stirred for 10 min at room temperature. The reaction was quenched by adding a solution of phosphate buffer (pH 5) and extracted with EtOAc. The combined organic layers were dried with Na₂SO₄ and evaporated; the mixture was subjected to column chromatography (silica gel; petroleum ether/EtOAc, 3:7) to give compound **24** (40 mg, 0.14 mmol, 61%). Colourless oil. $[α]_D^{24} = -26.79$ ($c = 0.74$, CH₃COCH₃). IR (film): $\tilde{\nu} = 3403, 1772, 1653, 1267, 1146$ cm⁻¹. ¹H NMR (300 MHz, CDCl₃/MeOD, 9:1): $\delta = 2.72$ (dd, $J = 11.1, 2.4$ Hz, 1 H, 1-H), 2.06 (m, 2 H, 2-H^A), 1.39 (m, 1 H, 2-H^B), 2.45 (ddd, $J = 12.9, 5.7, 3.0$ Hz, 1 H, 3-H^A), 2.25 (m, 1

H, 3-H^B), 5.31 (d, $J = 9.3$ Hz, 1 H, 5-H), 4.45 (dd, $J = 9.3, 9.3$ Hz, 1 H, 6-H), 2.82 (dddd, $J = 9.3, 6.6, 3.0, 3.0$ Hz, 1 H, 7-H), 4.08 (br. dd, $J = 9.3, 6.6$ Hz, 1 H, 8-H), 2.61 (br. d, $J = 14.1$ Hz, 1 H, 9-H^A), 1.50 (dd, $J = 14.1, 9.6$ Hz, 1 H, 9-H^B), 6.31 (d, $J = 3.0$ Hz, 1 H, 13-H^A), 6.26 (d, $J = 3.0$ Hz, 1 H, 13-H^B), 1.22 (s, 3 H, 14-H₃), 4.27 (d, $J = 12.9$ Hz, 1 H, 15-H^A), 3.93 (d, $J = 12.9$ Hz, 1 H, 15-H^B) ppm. ¹³C NMR (75.3 MHz, CDCl₃/MeOD, 9:1): see Table 2. MS (ESI⁺): m/z (%) = 319 (24) [M + K]⁺, 303 (100) [M + Na]⁺. C₁₅H₂₀O₅ (280.1311): calcd. C 64.27, H 7.19; found C 64.29, H 7.15.

(1R,4S,5S,6R,7S,8S,10R)-1,6-Dihydroxy-4(15)-epoxyeudesma-11(13)-en-8,12-olide (25) and (1R,4S,5S,6R,7S,8S,10R)-1,6-Dihydroxy-15-oxoeudesma-11(13)-en-8,12-olide (26): A solution of CHCl₃ saturated with gaseous HCl (1.5 mL) was added to a solution of **24** (40 mg, 0.14 mmol) in CHCl₃ (4 mL), whilst stirring. After 4 h, the solvent was evaporated and the mixture was subjected to column chromatography (silica gel; DCM/MeOH, 24:1) to give compounds **25** (5 mg, 0.02 mmol, 14%) and **26** (11 mg, 0.04 mmol, 29%). Data for **25**: Colourless oil. $[α]_D^{24} = -122.43$ ($c = 0.07$, CHCl₃). IR (film): $\tilde{\nu} = 3420, 1632$ cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 3.59$ (dd, $J = 10.5, 5.0$ Hz, 1 H, 1-H), 2.01 (m, 1 H, 2-H^A), 1.81 (m, 1 H, 2-H^B), 2.11 (m, 1 H, 3-H^A), 1.39 (ddd, $J = 13.2, 4.8, 2.4$ Hz, 1 H, 3-H^B), 1.86 (d, $J = 9.6$ Hz, 1 H, 5-H), 3.85 (dd, $J = 9.6, 9.6$ Hz, 1 H, 6-H), 2.54 (dddd, $J = 11.7, 9.6, 3.0, 3.0$ Hz, 1 H, 7-H), 3.96 (ddd, $J = 11.7, 11.7, 3.6$ Hz, 1 H, 8-H), 2.49 (dd, $J = 11.7, 3.6$ Hz, 1 H, 9-H^A), 1.55 (dd, $J = 11.7, 11.7$ Hz, 1 H, 9-H^B), 6.15 (d, $J = 3.0$ Hz, 1 H, 13-H^A), 5.97 (d, $J = 3.0$ Hz, 1 H, 13-H^B), 1.00 (s, 3 H, 14-H₃), 3.20 (dd, $J = 3.6, 1.2$ Hz, 1 H, 15-H^A), 2.86 (d, $J = 3.6$ Hz, 1 H, 15-H^B) ppm. ¹³C NMR (75.3 MHz, CDCl₃): see Table 2. MS (ESI⁺): m/z (%) = 319 (28) [M + K]⁺, 303 (100) [M + Na]⁺. C₁₅H₂₀O₅ (280.1311): calcd. C 64.27, H 7.19; found C 64.30, H 7.19. Data for **26**: Colourless oil. $[α]_D^{24} = -8.58$ ($c = 0.25$, CH₃OH). IR (film): $\tilde{\nu} = 3393, 1631, 1248, 992$ cm⁻¹. ¹H NMR (300 MHz, CDCl₃/MeOD, 9:1): $\delta = 3.24$ (dd, $J = 10.8, 5.1$ Hz, 1 H, 1-H), 1.62 (o.s., 1 H, 2-H^A), 1.53 (m, 1 H, 2-H^B), 2.31 (m, 1 H, 3-H^A), 1.36 (o.s., 1 H, 3-H^B), 2.87 (m, 1 H, 4-H), 1.60 (dd, $J = 10.2, 5.4$ Hz, 1 H, 5-H), 4.26 (dd, $J = 10.2, 10.2$ Hz, 1 H, 6-H), 2.45 (dddd, $J = 12.2, 10.2, 3.0, 3.0$ Hz, 1 H, 7-H), 3.91 (ddd, $J = 12.2, 11.7, 3.7$ Hz, 1 H, 8-H), 2.37 (dd, $J = 11.7, 3.7$ Hz, 1 H, 9-H^A), 1.36 (dd, $J = 11.7, 11.7$ Hz, 1 H, 9-H^B), 6.08 (d, $J = 3.0$ Hz, 1 H, 13-H^A), 5.94 (d, $J = 3.0$ Hz, 1 H, 13-H^B), 0.72 (s, 3 H, 14-H₃), 9.85 (br. s, 1 H, 15-H) ppm. ¹³C NMR (75.3 MHz, CDCl₃/MeOD, 9:1): see Table 2. MS (ESI⁺): m/z (%) = 319 (35) [M + K]⁺, 303 (100) [M + Na]⁺. C₁₅H₂₀O₅ (280.1311): calcd. C 64.27, H 7.19; found C 64.24, H 7.21.

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- [1] N. N. Fischer, E. J. Olivier, H. D. Fischer, *Prog. Chem. Org. Nat. Prod.* **1979**, 38, 77–390.
- [2] H. M. R. Hoffmann, J. Rabe, *Angew. Chem. Int. Ed. Engl.* **1985**, 24, 94–110.
- [3] B. M. Fraga, *Nat. Prod. Rep.* **2009**, 26, 1125–1155 and previous reviews.
- [4] A. A. S. Rodrigues, M. Garcia, J. A. Rabi, *Phytochemistry* **1978**, 17, 953–954.
- [5] F. S. El-Ferali, D. A. Benigni, A. T. McPhail, *J. Chem. Soc. Perkin Trans. 1* **1983**, 355–364.

- [6] T. Lu, N. H. Fischer, *Spectrosc. Lett.* **1996**, 29, 437–438.
- [7] A. F. Barrero, J. E. Oltra, V. Morales, M. Alvarez, I. Rodriguez-Garcia, *J. Nat. Prod.* **1997**, 60, 1034–1035.
- [8] T. R. Govindachari, B. S. Josi, V. N. Kamat, *Tetrahedron* **1965**, 21, 1509–1519.
- [9] T. A. Geissman, G. A. Ellestad, *Phytochemistry* **1971**, 10, 2475–2485.
- [10] T. S. Griffin, T. A. Geissman, T. W. Winters, *Phytochemistry* **1971**, 10, 2487–2495.
- [11] M. A. Irwin, K. H. Lee, R. F. Simpson, T. A. Geissman, *Phytochemistry* **1969**, 8, 2009–2012.
- [12] M. Bruno, S. Rosselli, A. Maggio, R. A. Raccuglia, K. F. Bastow, C. C. Wu, K. H. Lee, *Planta Med.* **2005**, 71, 1176–1178.
- [13] M. Bruno, S. Rosselli, A. Maggio, R. A. Raccuglia, K. F. Bastow, K.-H. Lee, *J. Nat. Prod.* **2005**, 68, 1042–1046.
- [14] S. Rosselli, A. Maggio, R. A. Raccuglia, M. Bruno, *Eur. J. Org. Chem.* **2003**, 2690–2694.
- [15] S. Rosselli, A. M. Maggio, R. A. Raccuglia, S. L. Morris-Natscke, K. F. Bastow, K. K. Lee, M. Bruno, *Nat. Prod. Commun.* **2010**, 5, 675–680.
- [16] M. Bruno, S. Rosselli, A. Maggio, R. A. Raccuglia, F. Senatore, F. Napolitano, *Planta Med.* **2003**, 63, 277–281.
- [17] A. F. Barrero, J. E. Oltra, I. Rodriguez, A. Barragan, D. G. Gravelos, P. Ruiz, *Fitoterapia* **1995**, 66, 227–230.
- [18] A. G. Gonzalez, V. Darias, G. Alonso, J. N. Boada, M. Fera, *Planta Med.* **1978**, 33, 356–359.
- [19] T. H. Porter, T. J. Mabry, H. Yoshioka, H. Fisher, *Phytochemistry* **1970**, 9, 199–204.
- [20] M. L. Jimeno, M. C. Aprea-Rojas, F. H. Cano, B. Rodriguez, *Magn. Reson. Chem.* **2004**, 42, 474–483.
- [21] J. E. Barquera-Lozada, G. Cuevas, *J. Org. Chem.* **2009**, 74, 874–883 and references cited therein.
- [22] M. Bruno, C. Fazio, M. P. Paternostro, J. G. Diaz, W. Herz, *Planta Med.* **1995**, 61, 374–375.
- [23] Yoshioka, W. Renold, T. J. Mabry, *J. Chem. Soc. C* **1970**, 148–149.
- [24] F. Bohlmann, J. Jakupovic, M. Ahmed, A. Schuster, *Phytochemistry* **1983**, 22, 1623–1636.
- [25] I. Merfort, G. Willuhn, D. Wendisch, D. Gondol, *Phytochemistry* **1996**, 42, 1093–1095.
- [26] F. Gao, H. Wang, T. J. Mabry, *Phytochemistry* **1990**, 29, 1601–1607.
- [27] F. Bohlmann, C. Zdero, H. Robinson, R. M. King, *Phytochemistry* **1980**, 19, 2473–2474.
- [28] A. F. Barrero, A. Rosales, J. M. Cuerva, J. E. Oltra, *Org. Lett.* **2003**, 5, 1935–1938.
- [29] M. C. de la Torre, B. Rodriguez, M. Bruno, C. Fazio, K. H. C. Baser, H. Dunam, *Phytochemistry* **1997**, 45, 1653–1662.
- [30] M. Bruno, S. Rosselli, A. Maggio, F. Piozzi, L. Scaglioni, O. Servettaz, *Biochem. Syst. Ecol.* **2004**, 32, 755–759.
- [31] V. Saroglou, A. Karioti, C. Demetozos, K. Dimas, H. Skaltsa, *J. Nat. Prod.* **2005**, 68, 1404–1407.
- [32] S. M. Adekenov, Yu. V. Gatilov, I. Yu. Bagryanskaya, V. A. Raldugin, *Sib. Khim. Zh.* **1993**, 76–79 [*Chem. Abstr.* 1993, **119**, 181139p].
- [33] J. A. Marco, J. F. Sanz-Cervera, V. Garcia-Lliso, L. R. Domingo, M. Carda, S. Rodriguez, F. Lopez-Ortiz, J. Lex, *Liebigs Ann.* **1995**, 1837–1841.
- [34] R. Azarken, F. M. Guerra, F. J. Moreno-Dorado, Z. D. Jorge, G. M. Massanet, *Tetrahedron* **2008**, 64, 10896–10905.
- [35] A. J. Minnaard, J. B. P. A. Wijnberg, A. de Groot, *J. Org. Chem.* **1997**, 62, 7346–7350.
- [36] A. Ortega, E. Maldonado, *Heterocycles* **1989**, 29, 635–638.

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